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Aflatoxin concentrations in locally processed Groundnut products in the Tamale Metropolis

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ABSTRACT

This study determined the aflatoxin levels in processed groundnut products in the Tamale metropolis. A simple random sampling technique was used to obtain roasted groundnuts, groundnut paste, kulikuli, and kulikulizim samples from three markets in the Metropolis. A total of 36 locally processed groundnut products were obtained and analyzed for total aflatoxin concentration, using the Reveal Q+ aflatoxin test method. All the groundnut products sampled tested positive with values ranging from 1.7 to 519 ppb. The most aflatoxin-contaminated product was kulikuli followed by kulikulizim. The roasted groundnut recorded the lowest mean total aflatoxin concentration of 21 ppb which was the closest to the acceptable limit of total aflatoxin (20 ppb) in Ghana. The high aflatoxin levels in kulikuli is a health concern that needs urgent attention.

Keywords: Aflatoxin; Kulikuli; Kulikulizim; Groundnut paste and Groundnut

INTRODUCTION

Aflatoxins are toxic secondary metabolites that are produced predominantly Aspergillus fungal species Aspergillus flavus, A. nomius, and A. parasiticus (Granados-Chinchilla et al., 2017; Juan et al., 2008; Shukla et al., 2017). They are low molecular weight (312 g/mol-328 g/mol) lipophilic compounds that are made up of alternating groups of ketone and methylene called polyketides which are carcinogenic and immunosuppressive to mammals (Amaike & Keller, 2011; Pandey et al., 2019; Sarmento Amoras & Pena Costa, 2020). Exposure to aflatoxins

through the consumption of contaminated foods may lead to acute poisoning (aflatoxicosis) which is characterized by a weakening immune system, impaired growth, and other health issues such as liver cancer in both livestock and humans (Alenyorege et al., 2015; Awuah & Kpodo, 1996; Coulombe et al., 2005; Hong et al., 2010). There are four naturally occurring aflatoxins which are B1 and B2 (produced by A. flavus), and G1 and G2 (produced by A. parasiticus). Among these, B1 is classified as a group 1, and the most potent carcinogen and genotoxic that affects all species mammalian even low concentrations (Pandey et al., 2019; Pitt,

The prevalence of aflatoxin has 2014). been reported in various food commodities from many parts of the world (Juan et al., 2008). Their contamination can occur at pre-harvest, harvest, and post-harvest stages in the food chain which is facilitated by various conditions including insect damage, grain physical injury, temperature, water activity, and humidity (Kebede et al., 2012; Marin et al., 2013). They have also been reported in processed foods such as cereal legume blends in Ghana (Opoku et. al., 2018). Aflatoxins affect the nutritional quality of products and cause economic loss to farmers and traders. Groundnut is one of the most susceptible substrates for aflatoxin production.

Groundnuts (Arachis hypogaea L.) also known as peanuts are one of the most important oilseed and food crops that are cultivated in the world (Prasad et al., 2010). Groundnuts can be used directly for food, or processed for cooking oil and a range of products (Shanahan et al., 2003; Singh & Singh, 1992). Due to its vitamins and mineral content groundnut products are utilized globally for the treatment and prevention of malnutrition in infants and young children, especially in less privileged homes that cannot afford animal protein (Achaglinkame et al., 2017; Asare et al., 2019; Masters et al., 2015). In Ghana, groundnuts are widely cultivated on small scale and consumed in almost every part of the country (Naab et al., 2005). It plays a very significant role in the lives of marginal farmers with respect to income and nutrition (Carlberg et al., 2014). In northern Ghana especially, groundnuts are eaten in roasted forms or processed into forms such kulikuli, paste, and kulikulizim (Florkowski & Kolavalli, 2013). The impact of aflatoxin contamination in groundnuts along the food chain has been a significant threat to trade, food security, and livelihoods in the country (Kumar et al., 2008). The stability of aflatoxin to varying processing conditions such as cooking, roasting, and extrusion makes it difficult to

eliminate it from contaminated food and food products. Therefore, maximum limits have been set in various countries for foods and feeds (García & Heredia, 2014). The Ghana Standards Authority, for instance, considers 10 ppb as the acceptable level of aflatoxin in groundnuts (Omari et al., 2022), while the European Union (EU), Codex Alimentarius Commission (Codex), and United States (USA) set 4 ppb, 15 ppb and 20 ppb, respectively, as the permissible limits (Agyekum & Jolly, 2017; Omari et al., 2022; WHO, 2004). Akullo et al. (2023) reported high levels of aflatoxin in groundnut products with 41.8% of the values higher than 20 ppb in the eastern and northern regions of Uganda. Similar reports have been made by different authors, in Zimbabwe (Dube & Maphosa, 2014), Nigeria (Ajeigbe et al., 2015), and Mali (Waliyar et al., 2015).

This study evaluated the occurrence of aflatoxins in groundnut-based products in the Tamale metropolis in the northern region of Ghana. The outcome of this study will help to create public awareness most especially for the safety and wellbeing of consumers.

MATERIALS AND METHODS

Sample collection

A simple random sampling technique was used to obtain samples from the markets. Three of each groundnut product was purchased from each market. A total of 36 groundnut products (roasted groundnut, groundnut paste, kulikuli, and kulikulizim) were purchased from 3 different markets (Aboabo, Central, and Lamashegu) in Tamale metropolis and stored in a refrigerator (at -20°C).

Sample preparation and extraction of groundnut samples

The samples were ground using a waring blender (HGBRWTS3-USA) and sieved using 20 mesh sieves. The sieved sample was extracted using 65% ethanol in distilled water with a sample-to-solvent ratio of 1:5. The resulting mixture was vigorously shaken in a well-covered glass bottle for 3 minutes. The mixture was then filtered with filter paper into a glass jar to obtain groundnut extract for analysis.

Total Aflatoxin Analysis

The samples were analyzed as described in the Reveal Q+ Test kit protocol for aflatoxin analysis. The Reveal Q+ method single-step is lateral flow immunochromatographic assay based on a competitive immunoassay format, designed for the quantification of aflatoxin in cereals, grain, and their products (Le et al., 2019). Briefly, the red sample dilution cups and white sample cups were placed into a sample cup rack and 500 µL of sample diluent was pipetted into each red sample solution cup. 100 µL of sample extract was then added to each red sample dilution cup containing the sample diluent. The mixing was done by pipetting up and down five times. Subsequently, 100 µL of each diluted sample extract was transferred into the white sample cups, and a new Reveal Q+ for Aflatoxin test strip was placed in the sample cup and a timer was set for 6 minutes to ensure wicking. After 6 minutes, it was removed and inserted into the cartridge. The cartridge with the test strip was then inserted into an mReader application one after the other. The mReader automatically analyzed the test strip and the results were displayed in part per billion (ppb).

Statistical Analysis

Data were entered into an EXCEL spreadsheet and analyzed by one-way analysis of variance (ANOVA) at a 5% level of significance using GenStat (12th edition) statistical software.

RESULTS AND DISCUSSION

Groundnut is a major dietary staple crop that is highly nutritious and high in energy owing to its high fat and protein content. It is widely cultivated and consumed in Ghana, Africa, and the world as a whole. However, it is one of the most susceptible crops to aflatoxin contamination (Jallow et al., 2021; Kumar et al., 2021). The impact of aflatoxin contamination along the food chain has been a major threat to trade, food security, and livelihoods. About two-thirds of farmers in the northern part of Ghana cultivate groundnuts on a relatively medium scale for sale, which is usually on a need basis and for home consumption. imperative Groundnut serves as an ingredient in local foods. Groundnut can be eaten roasted or processed into other forms such as paste which is used for soups and stews or into snacks such as kulikuli. Data on aflatoxin levels in locally processed groundnut products is limited in Ghana.

There was a highly significant difference (p = 0.004) within the locally processed groundnut products obtained from the markets. Kulikuli recorded the highest level of aflatoxin concentration (319 ppb), while roasted groundnut recorded the lowest (21 ppb) (Figure 1). The results showed that all the groundnut products had some levels of aflatoxin contamination. On average, only 17% of the products were below the various acceptable limits (4-20 ppb) for total aflatoxin in groundnut products. The high level of aflatoxin in three out of four of the products could be attributed to both primary secondary factors. Primary contamination arises from pre-harvest and post-harvest practices (Abass et al., 2014; Gachara et al., 2022; Mahuku et al., 2019) whereas secondary contamination normally occurs due to poor handling that allows inoculation of the product with the fungi during the production process (Chang et al., 2013).

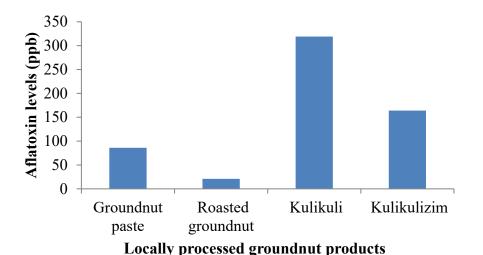


Figure 1: Average aflatoxin levels in various locally processed groundnut products

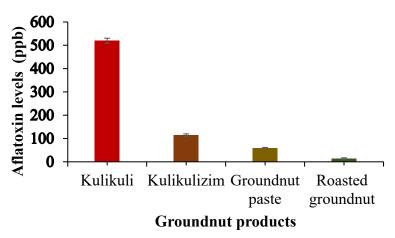


Figure 3: Average aflatoxin levels in the various locally processed groundnut products from the Central Market (error bars = SEM)

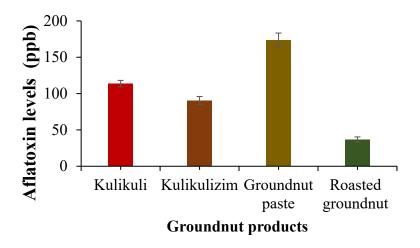


Figure 2. Aflatoxin level of processed groundnut product from Lamashegu market

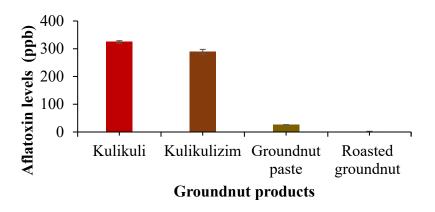


Figure 4: Average aflatoxin levels in the locally processed groundnut products in the Aboabo market (error bars = SEM)

Aflatoxin levels in locally processed groundnut products in various markets

Lamashegu market

In this market, groundnut paste had the highest mean value of 173 ppb, while roasted groundnut had the lowest aflatoxin concentration of 36 ppb (Figure 2).

Central Market and Aboabo Market

In the central market, kulikuli had the highest level of aflatoxin with a mean value of 519 ppb (Figure 3) while in the Aboabo market, kulikuli had the highest mean aflatoxin of 324 ppb. This was closely followed by kulikulizim (Figure 4).

Primarily, contamination may arise from pre-harvest and post-harvest practices (Abass et al., 2014; Gachara et al., 2022; Mahuku et al., 2019). Drought stress is one of the major contributing factors to precontamination harvest aflatoxin groundnuts (Guo et al., 2008) and this is exacerbated when coupled with high temperature and low atmospheric humidity (Achaglinkame et al., 2017; Molnár et al., 2023). Inappropriate post-harvest practices such as poor shelling can lead to damaged kernels which mechanically increases the ease of fungal attack and colonization thus and aflatoxin contamination. Poor storage is a critical postharvest practice that has the potential to increase aflatoxin buildup in groundnuts during storage. When nuts are not properly dried before storage or when there is reintroduction of moisture into nuts as a result of rewetting and activities such as respiration of rodents and insects, aflatoxin builds up rapidly.

The high level of aflatoxin in *kulikuli* can be attributed to the use of high-risk aflatoxin groundnuts. Currently in the Northern parts of Ghana, the practice among groundnut farmers and traders is

that, when bad nuts (high-risk aflatoxin groundnuts (HRAG)) are sorted from the lot, the good lot is sold and the HRAG are either mixed with relatively good ones and consumed by the household or sold at low prices to retailers. The HRAGs thus find their way back into the food value chain when producers of kulikuli and kulikulizim purchase them due to their low price, and mix with relatively better nuts but of higher production price, to reduce production cost. HRAG, normally, would have a rancid to bitter taste. However, when used in relatively low amounts in products like kulikuli, kulikulizim, and groundnut paste, the bitterness is masked, most especially in kulikuli and kulikulizim due to the addition of spices to these two products. This may explain the higher aflatoxin levels in kulikuli and kulikulizim. Again, kulikuli, kulikulizim and groundnut paste are all milled in commercial milling machines during processing.

In all three markets, roasted groundnuts consistently recorded lower aflatoxin levels compared to the other groundnut products. Roasted groundnut, is normally processed by sorting before roasting. Sorting is known to reduce aflatoxin by about 40-80% (Park, 2002). Thus, the low levels of aflatoxin in roasted groundnuts from all the markets suggest that sorting was done before roasting. This is understandable because a single HRAG can easily be noticed in packaged roasted nuts or taste bad once chewed on, which can lead to reduced acceptability by consumers.

CONCLUSION

This study evaluated the aflatoxin concentration in groundnut products. Out of the 36 groundnut products collected from the three markets, only 17% which happened to be roasted groundnut samples had aflatoxin levels (1.7 -20 ppb) within the permissible limits in Ghana. The extremely

high levels of kulikuli raise serious health concerns especially when it is used as a snack for children. This calls for education and sensitization of stakeholders on the necessary value chain management approach in reducing aflatoxin levels. The efforts by food processors and vendors in controlling aflatoxin levels and measures by farmers to control pre- and post-harvest contamination are essential to ensure the quality of these products.

Conflicts of interest

The authors declare that there are no conflicts of interest.

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REFERENCES

- Abass, A. B., Ndunguru, G., Mamiro, P., Alenkhe, B., Mlingi, N., & Bekunda, M. (2014). Post-harvest food losses in a maize-based farming system of semi-arid savannah area of Tanzania. *Journal of Stored Products Research*, 57, 49-57.
- Achaglinkame, M. A., Opoku, N., & Amagloh, F. K. (2017). Aflatoxin contamination in cereals and legumes to reconsider usage as complementary food ingredients for Ghanaian infants: A review.

- Journal of nutrition & intermediary metabolism, 10, 1-7.
- Agyekum, M., & Jolly, C. M. (2017).

 Peanut trade and aflatoxin standards in Europe: Economic effects on trading countries. *Journal of Policy Modeling*, 39(1), 114-128.

 https://doi.org/https://doi.org/10.10
 16/j.jpolmod.2016.08.004
- Ajeigbe, H. A., Waliyar, F., Echekwu, C., Kunihya, A., Motagi, B., Eniaiyeju, D., & Inuwa, A. (2015). A farmer's guide to profitable groundnut production in Nigeria.
- Akullo, J. O., Amayo, R., Okello, D. K., Mohammed, A., Muyinda, R., Magumba, D., Gidoi, R., & Mweetwa, A. M. (2023). Aflatoxin contamination in groundnut and maize food products in Eastern and Northern Uganda. *Cogent Food & Agriculture*, 9(1), 2221015.
- Alenyorege, E. A., Abagale, F. K., Yawson, A., & Opoku, N. (2015). Effects of Fertilization on Aflatoxin Concentration in Fresh and Stored Groundnuts (*Arachis hypogaea L.*). *International Journal of Science, Engineering and Technology*, 3165-3169.
- Amaike, S., & Keller, N. P. (2011). Aspergillus flavus. *Annual review of phytopathology, 49*(1), 107-133.
- Asare B. K., Ofori, K., Offei, S. K., Dzidzienyo, D., Asibuo, J. Y., & Adu Amoah, R. (2019). Aflatoxin contamination of groundnut (Arachis hypogaea L.): Predisposing factors and management interventions. Food Control, 98, 61-67. https://doi.org/https://doi.org/10.10 16/j.foodcont.2018.11.020

- Awuah, R. T., & Kpodo, K. A. (1996). High incidence of Aspergillus flavus and aflatoxins in stored groundnut in Ghana and the use of a microbial assay to assess the inhibitory effects of plant extracts on aflatoxin synthesis. *Mycopathologia*, *134*(2), 109-114. https://doi.org/10.1007/bf00436873
- Carlberg, E., Kostandini, G., & Dankyi, A. (2014). The Effects of Integrated Pest Management Techniques Farmer Field Schools on Groundnut Productivity: Evidence from Ghana. *Quarterly Journal of International Agriculture*, 53(1), 73-88.
- Chang, A. S., Sreedharan, A., & Schneider, K. R. (2013). Peanut and peanut products: a food safety perspective. *Food Control*, 32(1), 296-303.
- Coulombe, R. A., Guarisco, J. A., Klein, P. J., & Hall, J. O. (2005). Chemoprevention of aflatoxicosis in poultry by dietary butylated hydroxytoluene. *Animal Feed Science and Technology, 121*(1), 217-225. https://doi.org/https://doi.org/10.1016/j.anifeedsci.2005.03.001
- Dube, M., & Maphosa, M. (2014).

 Prevalence of aflatoxigenic
 Aspergillus spp and groundnut
 resistance in Zimbabwe. *J. Agric.*Vet. Sci, 7, 8-12.
- Florkowski, W. J., & Kolavalli, S. (2013). Aflatoxin control strategies in the groundnut value chain in Ghana. *IFPRI Ghana Strategy Support Program Working Paper*, 33.
- Gachara, G., Suleiman, R., El Kadili, S., Ait Barka, E., Kilima, B., & Lahlali, R. (2022). Drivers of post-harvest aflatoxin contamination: evidence gathered from knowledge

- disparities and field surveys of maize farmers in the rift valley region of Kenya. *Toxins*, 14(9), 618.
- García, S., & Heredia, N. L. (2014).
 Aflatoxins: An Overview. Aflatoxin
 Control: Safeguarding Animal Feed
 with
 Calcium
 Smectite(aflatoxincontro), 1-10.
- Granados-Chinchilla, F., Molina, A., Chavarría, G., Alfaro-Cascante, M., Bogantes-Ledezma, D., & Murillo-Williams, A. (2017). Aflatoxins occurrence through the food chain in Costa Rica: Applying the One Health approach to mycotoxin surveillance. Food Control, 82, 217-226.
 - https://doi.org/https://doi.org/10.10 16/j.foodcont.2017.06.023
- Guo, B., Chen, X., Dang, P., Scully, B. T., Liang, X., Holbrook, C. C., Yu, J., & Culbreath, A. K. (2008). Peanut gene expression profiling developing seeds at different reproduction during stages Aspergillus parasiticus infection. BMC Developmental Biology, 8(1), 12.
- Hong, L. S., Mohd Yusof, N., & Ling, H. M. (2010). Determination of aflatoxins B1 and B2 in peanuts and corn based products. *Sains Malaysiana*, 39(5), 731-735.
- Jallow, A., Xie, H., Tang, X., Qi, Z., & Li, P. (2021). Worldwide aflatoxin contamination of agricultural products and foods: From occurrence to control. Comprehensive Reviews in Food Science and Food Safety, 20(3), 2332-2381. https://doi.org/https://doi.org/10.11
- Juan, C., Zinedine, A., Moltó, J. C., Idrissi, L., & Mañes, J. (2008). Aflatoxins

11/1541-4337.12734

- levels in dried fruits and nuts from Rabat-Salé area, Morocco. *Food Control*, 19(9), 849-853. https://doi.org/https://doi.org/10.10 16/j.foodcont.2007.08.010
- Kebede, H., Abbas, H. K., Fisher, D. K., & Bellaloui, N. (2012). Relationship between aflatoxin contamination and physiological responses of corn plants under drought and heat stress. *Toxins*, 4(11), 1385-1403.
- Kumar, A., Pathak, H., Bhadauria, S., & Sudan. J. (2021).Aflatoxin contamination in food crops: causes, detection, and management: review. Food Production, Processing and Nutrition, 3(1), 17. https://doi.org/10.1186/s43014-021-00064-y
- Kumar, V., Basu, M. S., & Rajendran, T. P. (2008). Mycotoxin research and mycoflora in some commercially important agricultural commodities. *Crop protection*, 27(6), 891-905. https://doi.org/http://dx.doi.org/10.1016/j.cropro.2007.12.011
- Le, O.-N., Roman, B., Driksna, D., Gilbert, L., Gonzales, K., Klein, Donofrio, R., Shephard, G., Trucksess, M., & Ziemer, W. (2019). Reveal Q+ MAX® for Detection of Total Af latoxin in Corn. Almonds, Pistachios. Walnuts, and Peanuts. Journal of AOAC International, 102(2), 525-531.
- Mahuku, G., Nzioki, H. S., Mutegi, C., Kanampiu, F., Narrod, C., & Makumbi, D. (2019). Pre-harvest management is a critical practice for minimizing aflatoxin contamination of maize. *Food Control*, *96*, 219-226.
 - https://doi.org/https://doi.org/10.10 16/j.foodcont.2018.08.032

- Marin, S., Ramos, A. J., Cano-Sancho, G., & Sanchis, V. (2013). Mycotoxins: Occurrence, toxicology, and exposure assessment. *Food and Chemical Toxicology*, 60, 218-237. https://doi.org/http://dx.doi.org/10.1016/j.fct.2013.07.047
- Masters, W. A., Ghosh, S., Daniels, J. A., & Sarpong, D. B. (2015). Comprehensive assessment of the peanut value chain for nutrition improvement in Ghana final report, september 2013. Accessed December, 28.
- Molnár, K., Rácz, C., Dövényi-Nagy, T., Bakó, K., Pusztahelyi, T., Kovács, S., Adácsi, C., Pócsi, I., & Dobos, A. (2023). The effect of environmental factors on mould counts and AFB1 toxin production by Aspergillus flavus in maize. *Toxins*, 15(3), 227.
- Naab, J., Tsigbey, F., Prasad, P., Boote, K., Bailey, J., & Brandenburg, R. (2005). Effects of sowing date and fungicide application on yield of early and late maturing peanut cultivars grown under rainfed conditions in Ghana. *Crop protection*, 24(4), 325-332.
- Omari, R., M. Akuffobea-Essilfie, E. K. Tetteh, N. Y. Asafu-Adjaye, E. Jumpah, and I. B. Gokah. 2022. Prioritizing Ghana's Aflatoxin Policy Implementation Plan using P-IMA. https://standardsfacility.org/sites/default/files/Prioritizing_Ghana_Aflatoxin_Policy_Implementation_ Plan_P-IMA%20-%20Final.pdf
- Opoku N, Achaglinkame M A and Amagloh F K. 2018. Aflatoxin content in cereal-legume blends on the Ghanaian market far exceeds the permissible limits. *Food security*.10:6, 1539-1545

- WHO (2004). Food and agriculture organization of the United Nations. *Vitamin and mineral requirements in human nutrition*, *2*, 17-299.
- Pandey, M. K., Kumar, R., Pandey, A. K., Soni, P., Gangurde, S. S., Sudini, H. K., Fountain, J. C., Liao, B., Desmae, H., & Okori, P. (2019). Mitigating aflatoxin contamination in groundnut through a combination of genetic resistance and post-harvest management practices. *Toxins*, 11(6), 315.
- Park, D. L. (2002). Effect of processing on aflatoxin. *Mycotoxins and Food Safety*, 173-179.
- Pitt, J. I. (2014). Mycotoxins: Aflatoxins. In Y. Motarjemi (Ed.), *Encyclopedia of Food Safety* (pp. 289-294). Academic Press. https://doi.org/https://doi.org/10.1016/B978-0-12-378612-8.00190-6
- Prasad, P. V., Kakani, V. G., & Upadhyaya, H. D. (2010). Growth and production of groundnut. *UNESCO Encyclopedia*, 1-26.
- Sarmento Amoras, E., & Pena Costa, A. L. (2020). Aflatoxins: A Brief Review of their Chemical Properties, Toxicological Effects and Control Measures.

- Shanahan, H., Carlsson-Kanyama, A., Offei-Ansah, C., Ekström, M. P., & Potapova, M. (2003). Family meals and disparities in global ecosystem dependency. Three examples: Ghana, Russia and Sweden. *International Journal of Consumer Studies*, 27(4), 283-293.
- Shukla, S., Kim, D. H., Chung, S. H., & Kim, M. (2017). Chapter 28 Occurrence of Aflatoxins in Fermented Food Products. In J. Frias, C. Martinez-Villaluenga, & E. Peñas (Eds.), Fermented Foods in Health and Disease Prevention (pp. 653-674). Academic Press. https://doi.org/https://doi.org/10.10 16/B978-0-12-802309-9.00028-5
- Singh, U., & Singh, B. (1992). Tropical grain legumes as important human foods. *Economic Botany*, 46(3), 310-321.
- Waliyar, F., Umeh, V., Traore, A., Osiru, M., Ntare, B., Diarra, B., Kodio, O., Kumar, K. V. K., & Sudini, H. (2015). Prevalence and distribution of aflatoxin contamination in groundnut (*Arachis hypogaea* L.) in Mali, West Africa. *Crop Protection*, 70, 1-7.